



FD-ISHB Research

Shannon Lynch and Akif Eskalen
Plant Pathology Department
UC Davis

Virtual Meeting of SDMMP Research Update
October 27, 2021



Acer negundo



Acacia melanoxylon



Koelreuteria elegans



Quercus agrifolia



Albizia kalkora



Platanus racemosa



Ficus sp.



Quercus robur



Persea americana



Populus fremontii



❖ Broad host range

❖ 77 species support beetle reproduction (competent)

❖ 20 native to California

❖ Avocado

❖ 25 – 60% percent trees in urban landscape



Platanus racemosa



Ficus sp.



Quercus robur

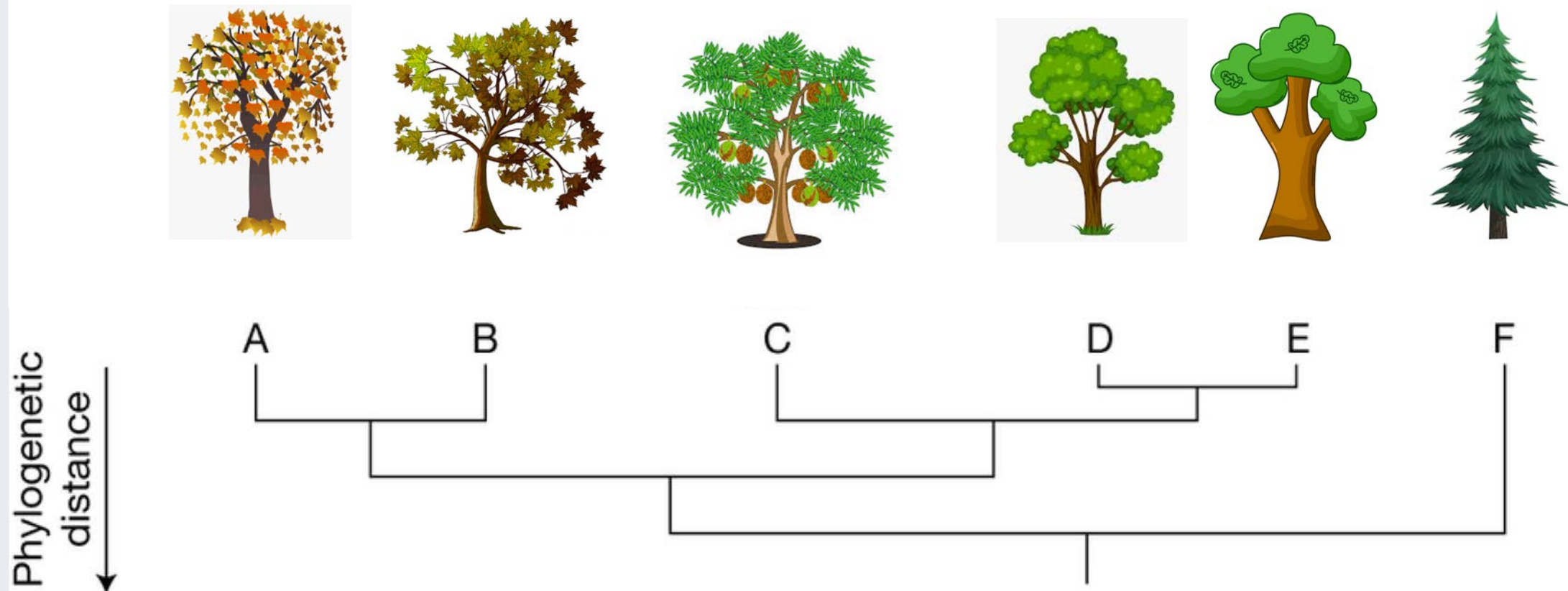


Persea americana

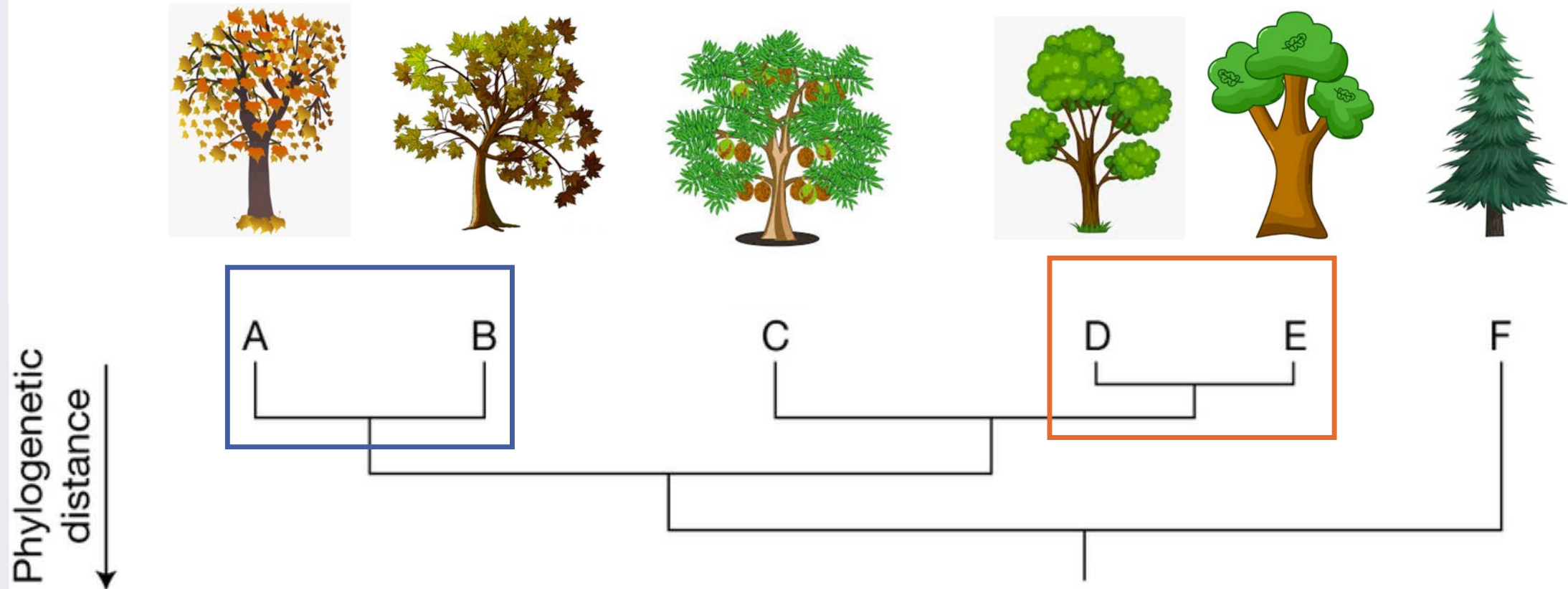


Populus fremontii

Phylogenetic Signal in Host Range



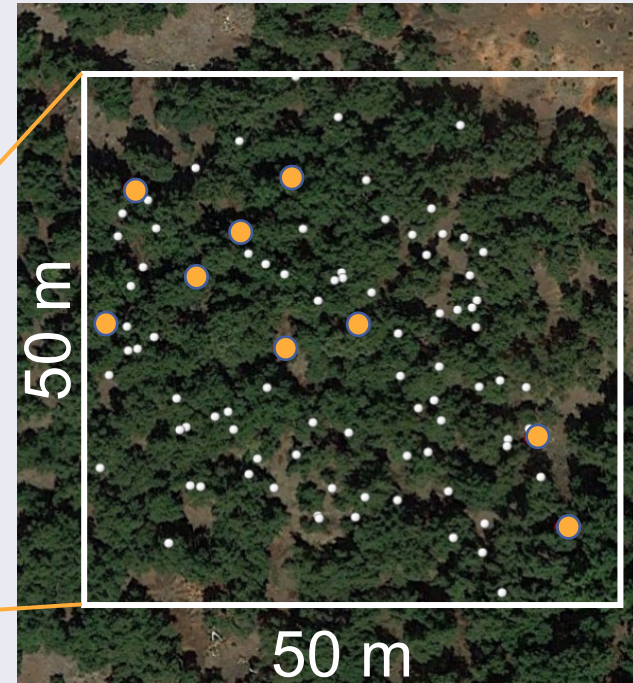
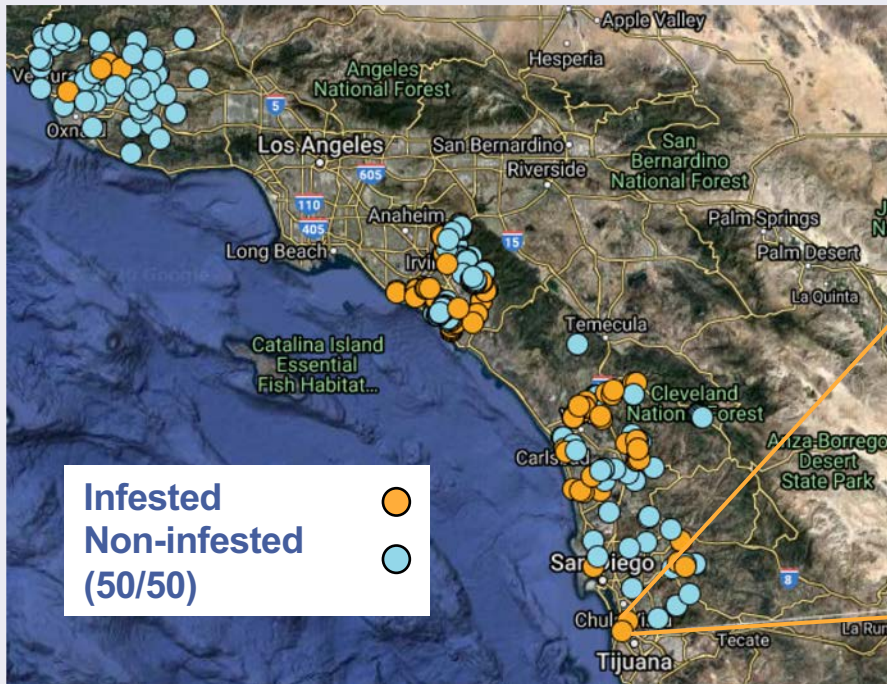
Phylogenetic Signal in Host Range



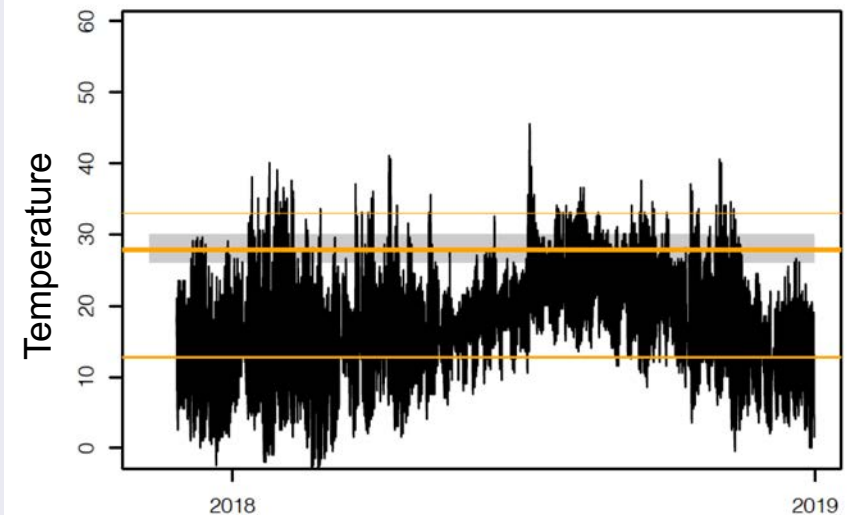
Predicting disease establishment in heterogeneous landscapes



Methods: Network of 260 Monitoring Plots



- 260 monitoring plots
- 83 in San Diego Co.
 - ❖ Host Composition
 - ❖ Microclimate
 - ❖ Disease Severity
 - ❖ Monitor annually since 2017

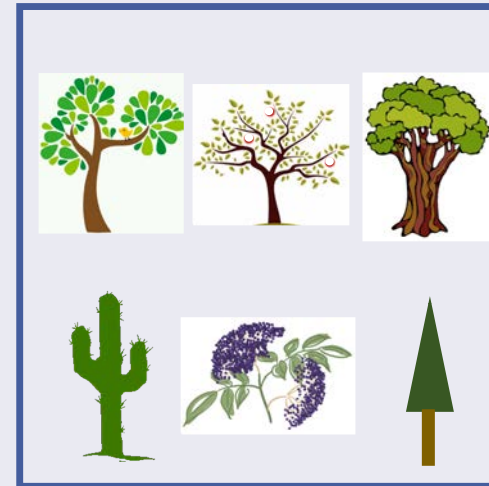


Predicting Disease Establishment

Host Abundance and Phylogenetic Structure



wpS =
Site
Susceptibility
Based on
Phylogenetic
Composition

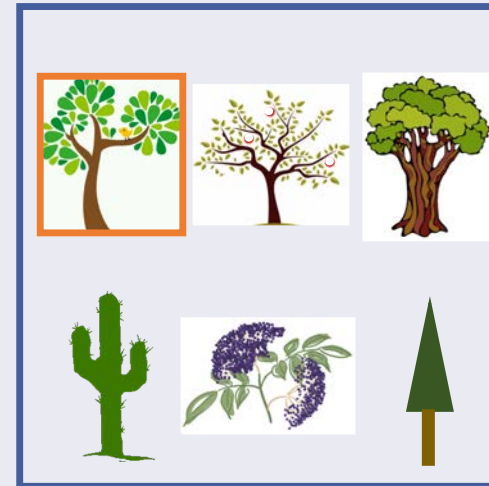


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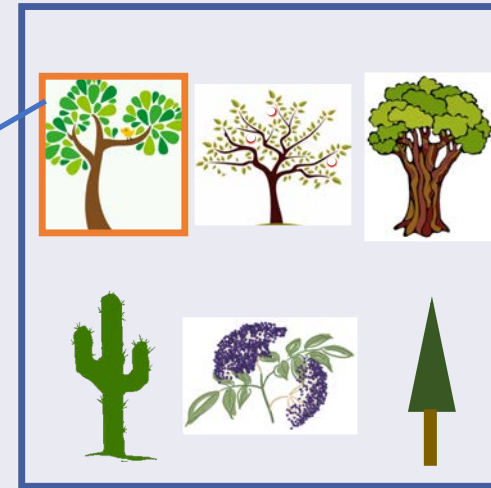
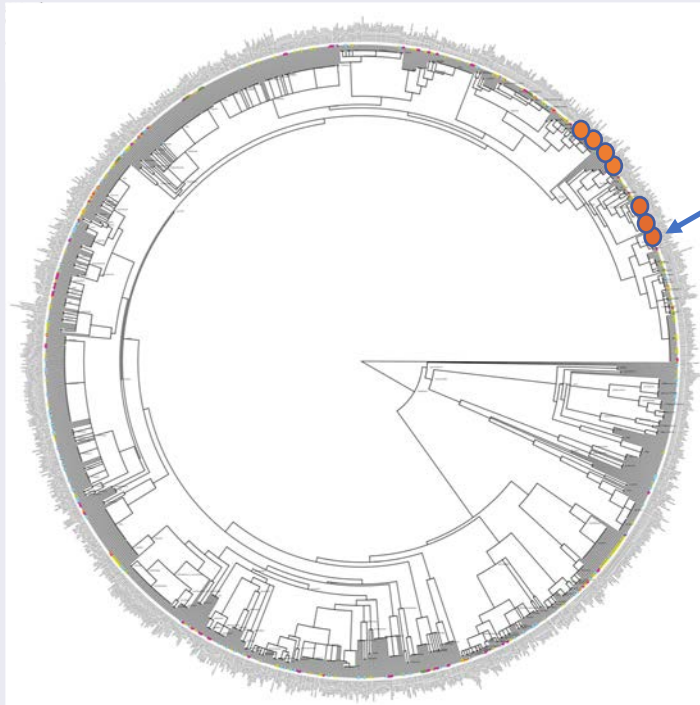


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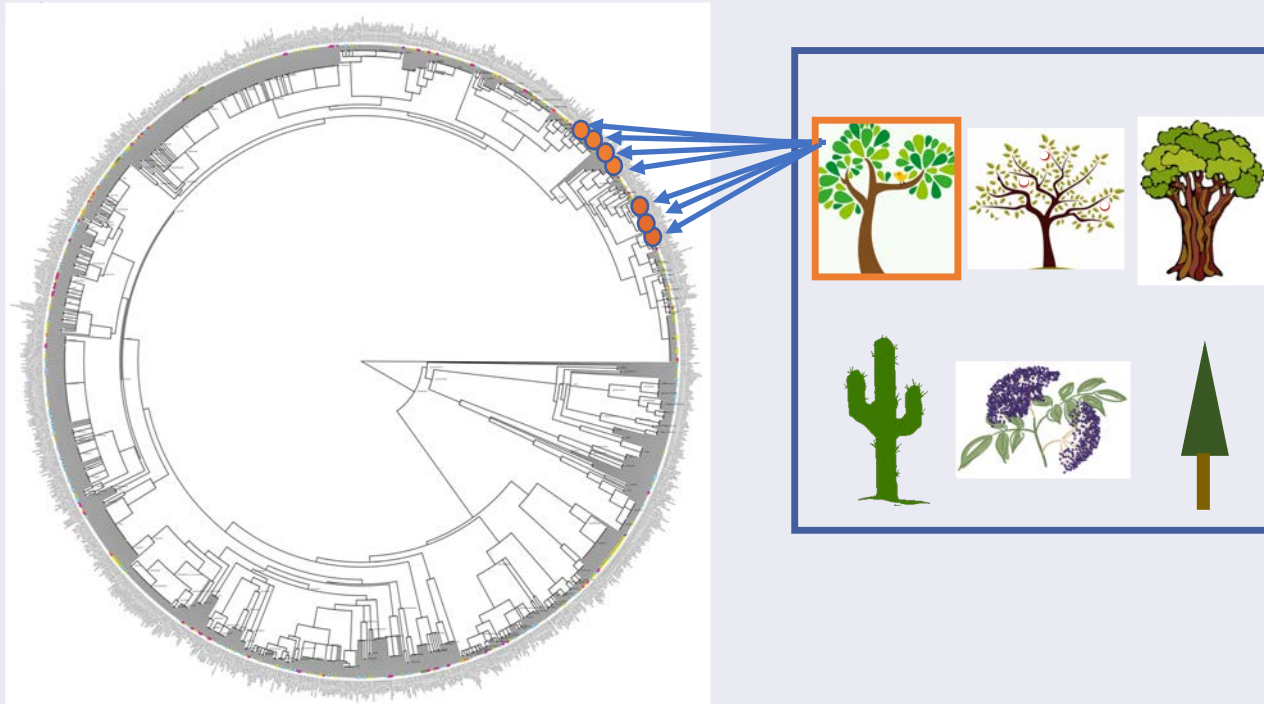


Predicting Disease Establishment

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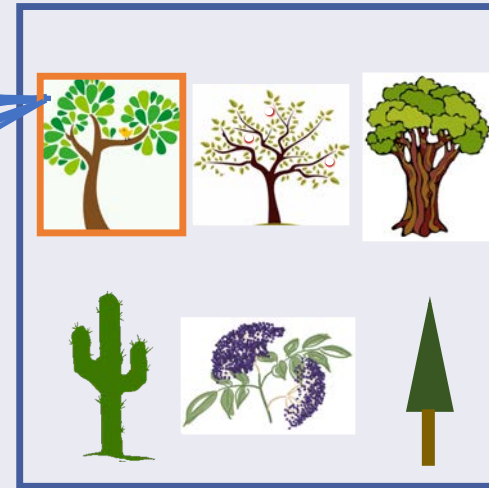
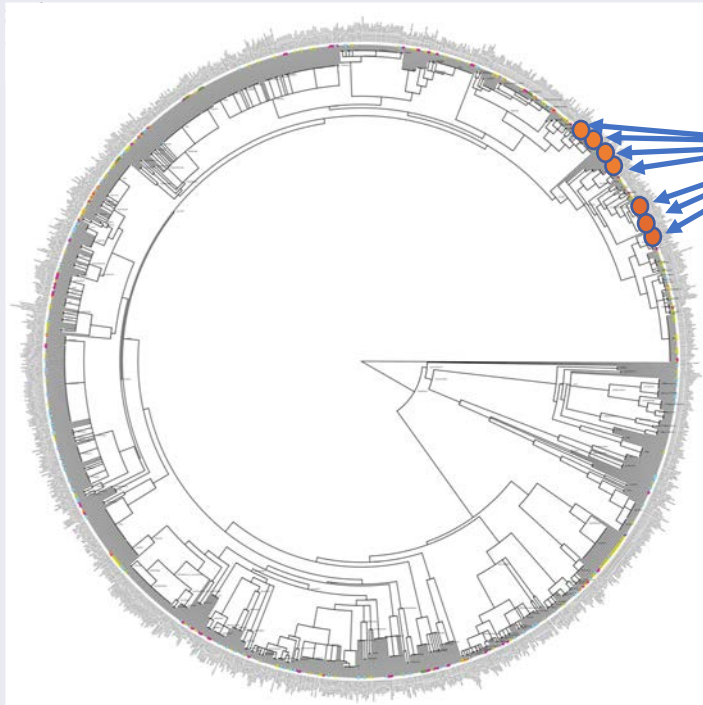


Predicting Disease Establishment

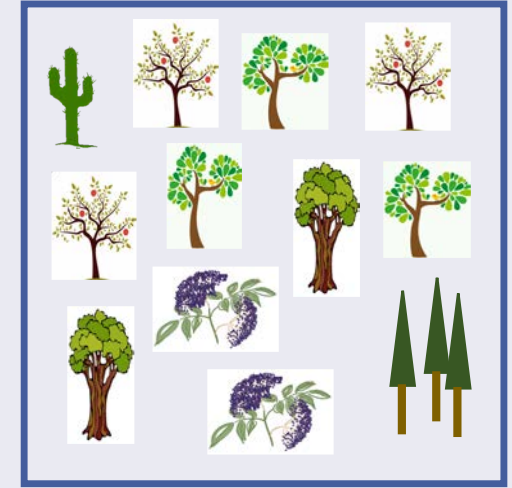
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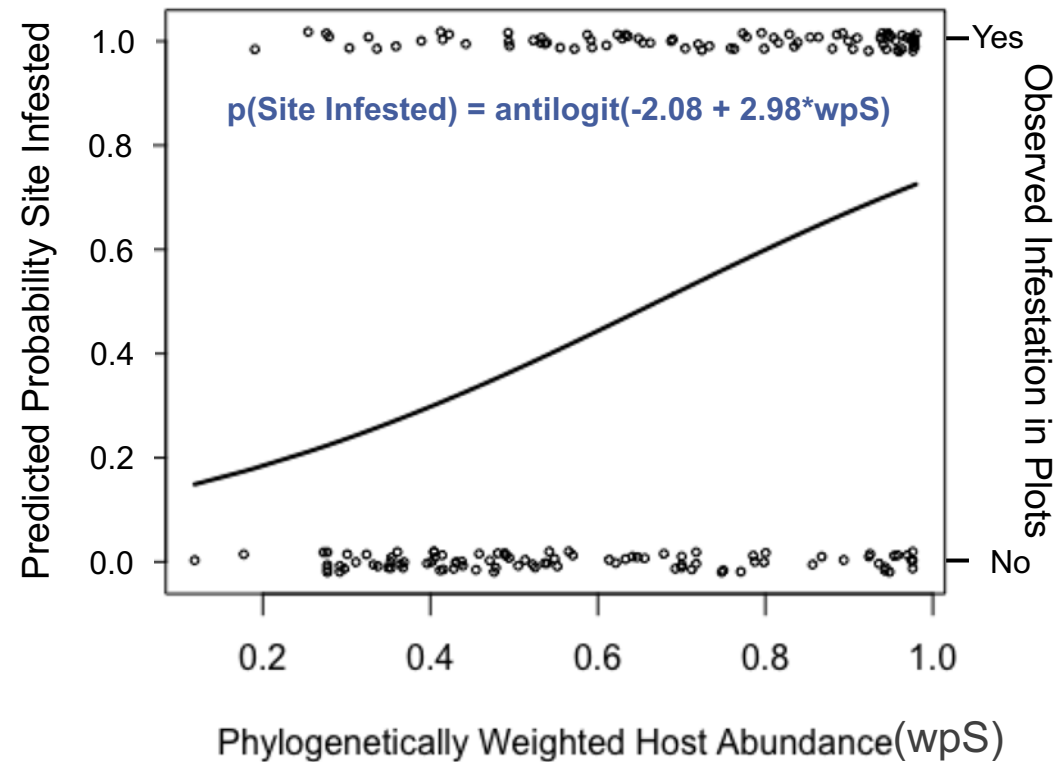
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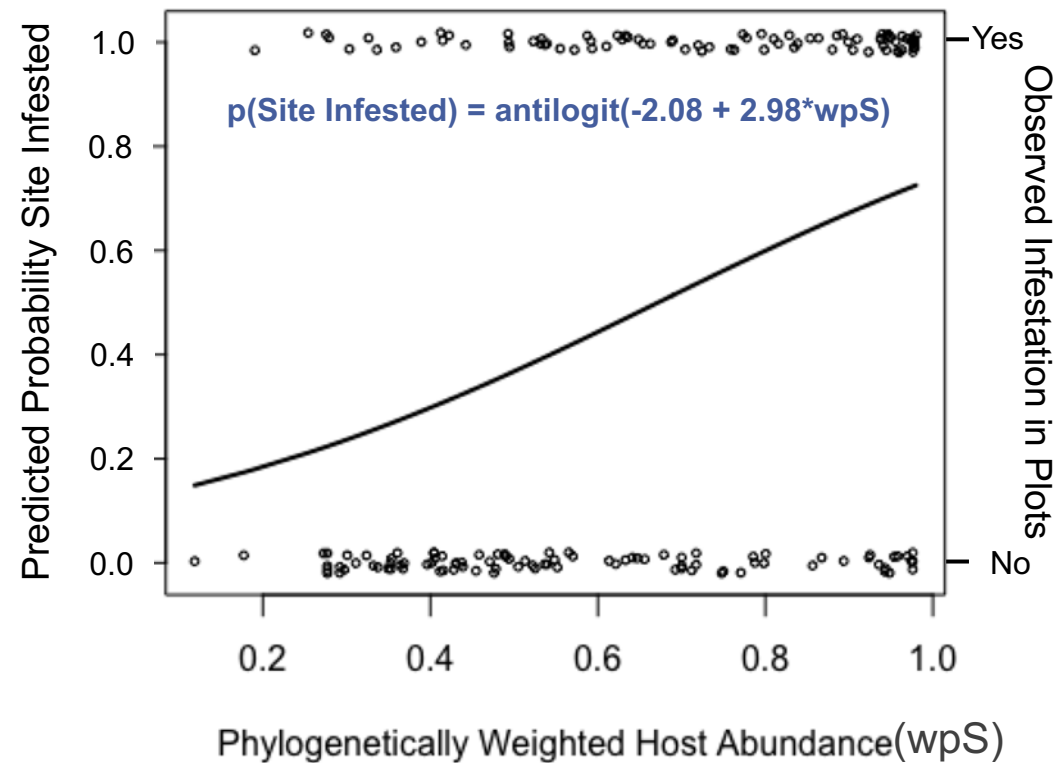


How well does wpS predict observed infestation in plots?

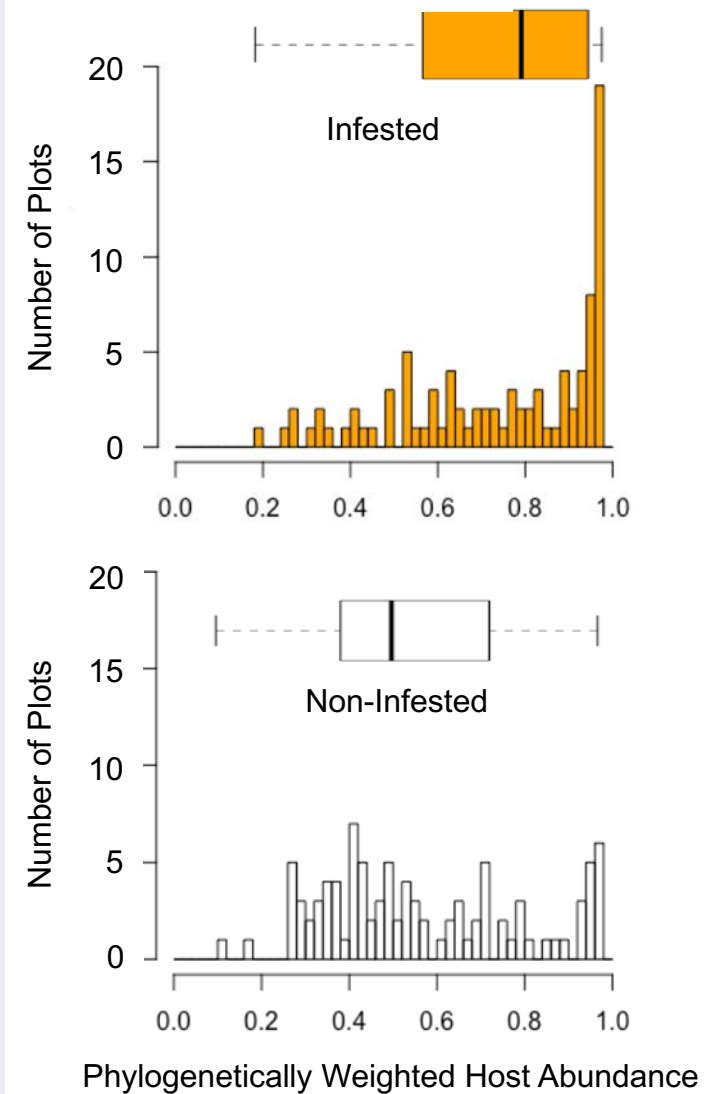


**Susceptibility Based on
Phylogenetic Composition**

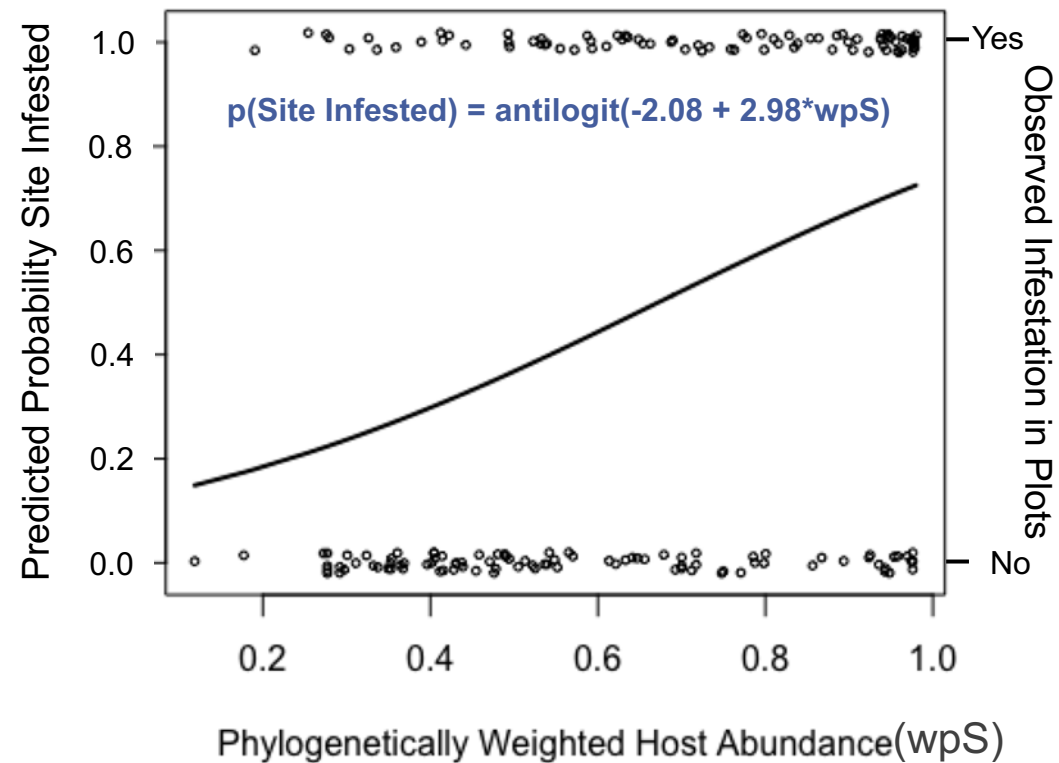
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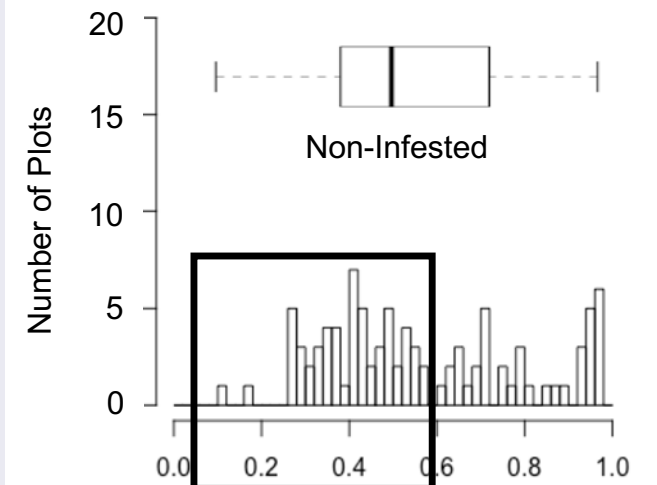
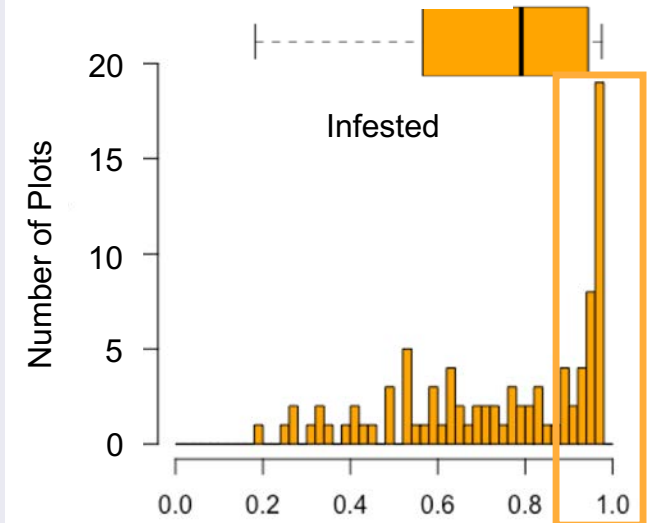
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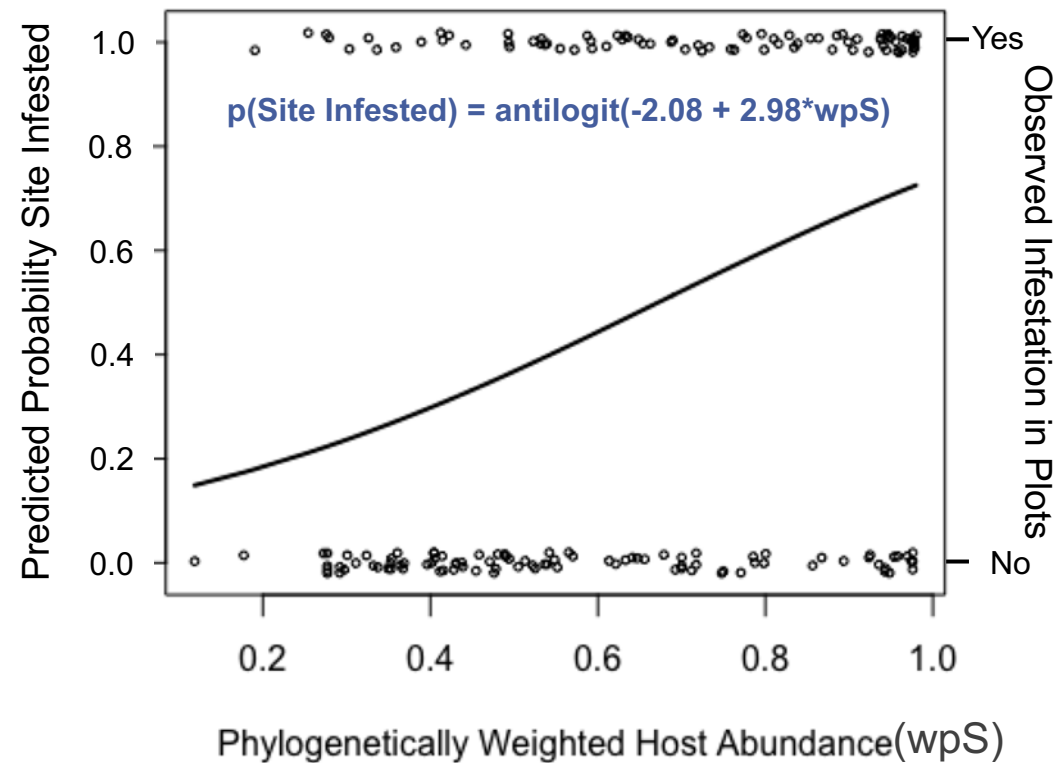


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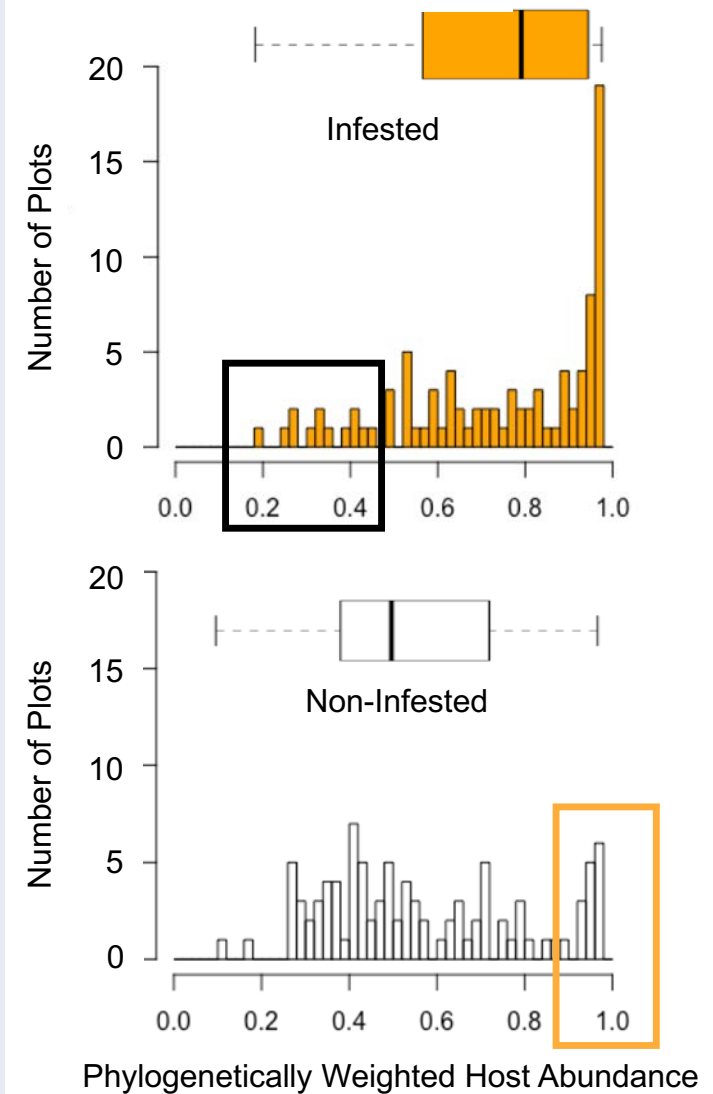


Phylogenetically Weighted Host Abundance

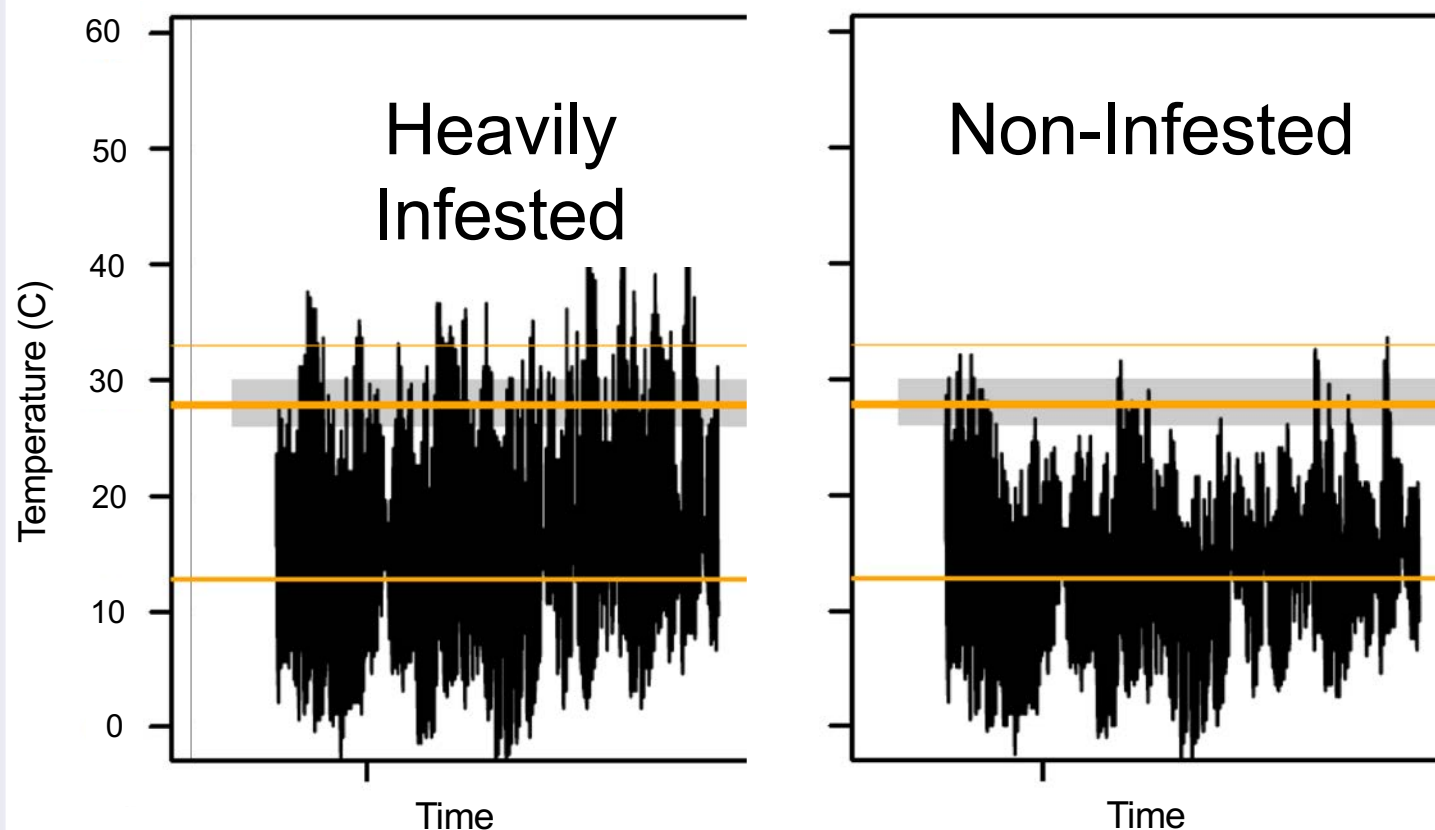
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**Susceptibility Based on
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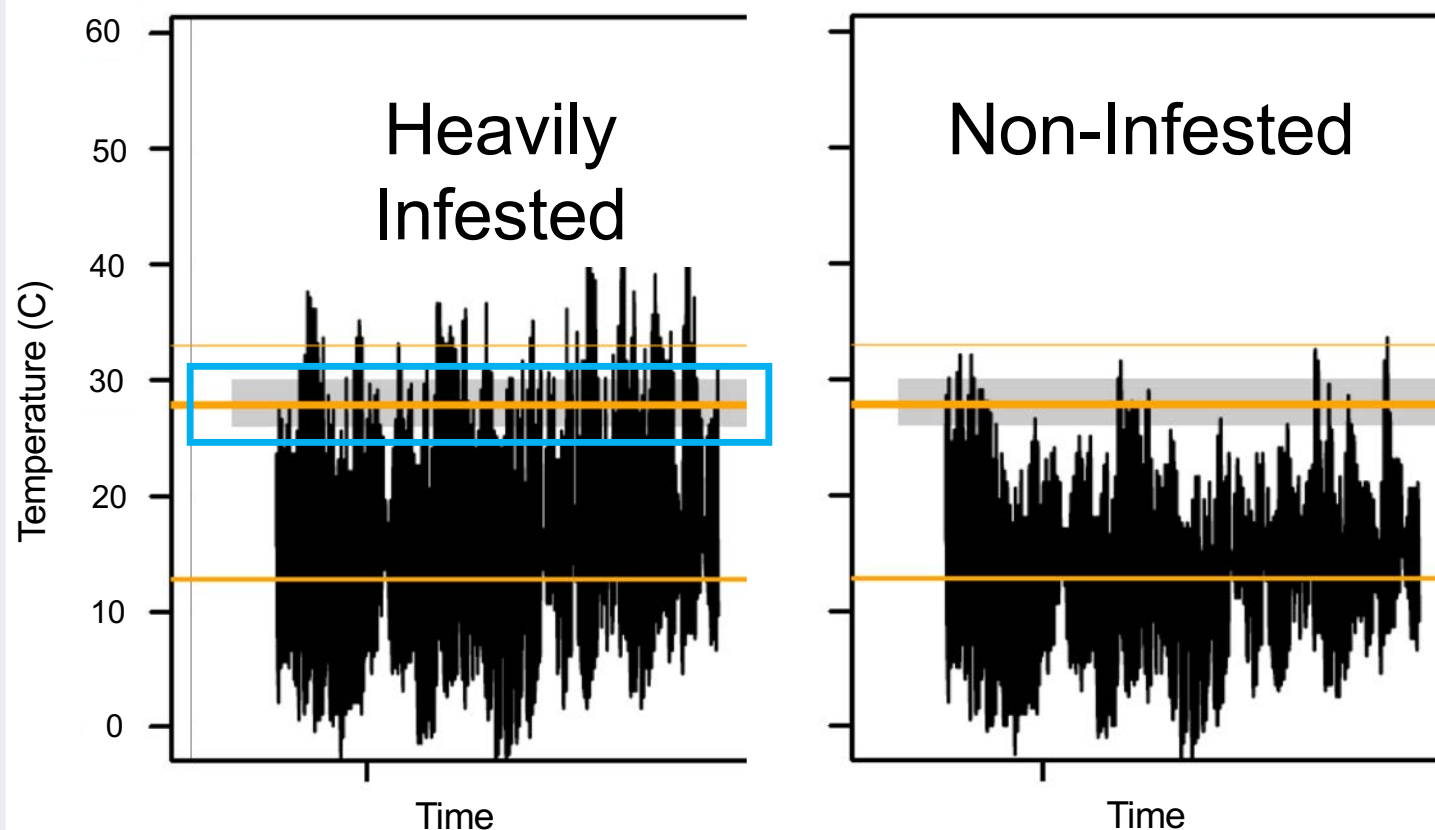


Degree Day Models and ISHB Development



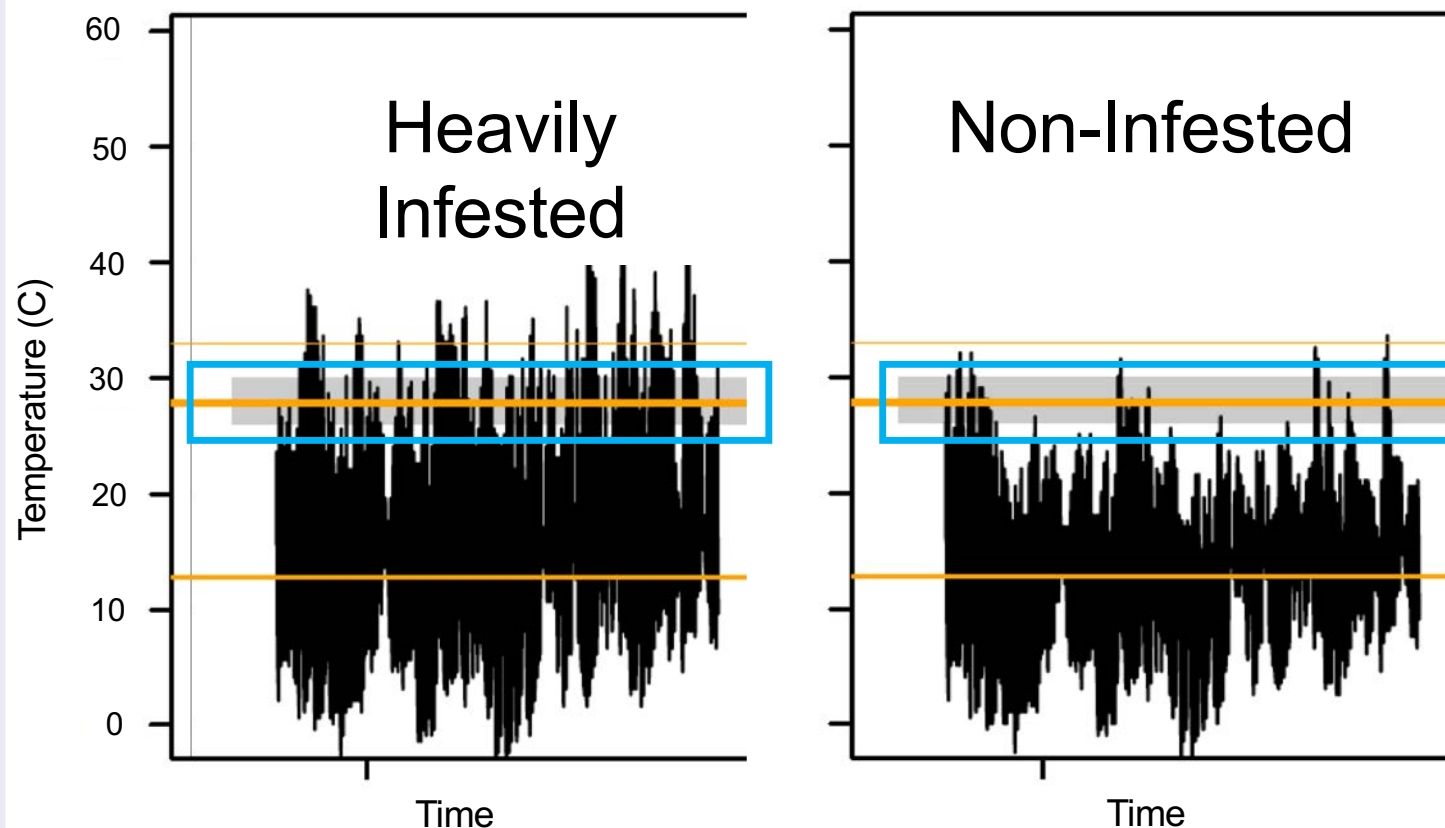
Species	T_{\min}	T_{opt}	T_{\max}	K	Reference
PSHB	13.34	27.51	33.08	398	Umeda & Paine 2018
KSHB	12.77	28.04	31.99	318	Dodge & Stouthamer (<i>in review</i>)

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Community Structure and Climate Effects



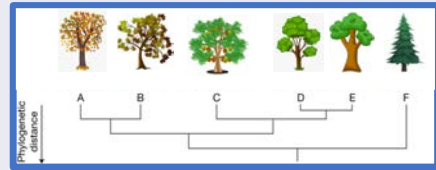
$$p(\text{Site Suscept}) = \left[\begin{array}{c} \text{Phylogenetic distance} \\ \downarrow \\ \text{A} \quad \text{B} \quad \text{C} \quad \text{D} \quad \text{E} \quad \text{F} \end{array} \right] + \# \text{ gens} \left[\begin{array}{c} \text{Temp (C)} \end{array} \right] + \left[\begin{array}{c} \text{Phylogenetic distance} \\ \downarrow \\ \text{A} \quad \text{B} \quad \text{C} \quad \text{D} \quad \text{E} \quad \text{F} \end{array} \right] * \# \text{ gens} \left[\begin{array}{c} \text{Temp (C)} \end{array} \right]$$

The diagram illustrates a conceptual model for site susceptibility. It consists of three main components arranged horizontally, separated by a plus sign and a multiplication sign. Each component features a phylogenetic tree with six tips labeled A through F, each associated with a different tree species illustration. The first component is labeled 'Phylogenetic distance' with a downward arrow. The second component is labeled 'Temp (C)' with a horizontal line. The third component is also labeled 'Phylogenetic distance' with a downward arrow. The entire expression is preceded by 'p(Site Suscept) ='.

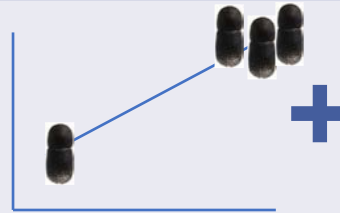
Community Structure and Climate Effects



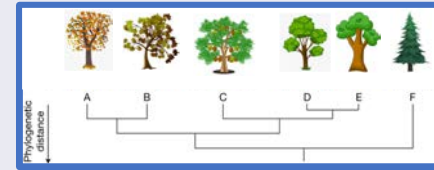
$$p(\text{Site Suscept}) =$$



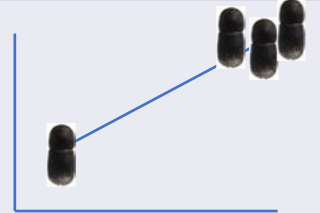
gens



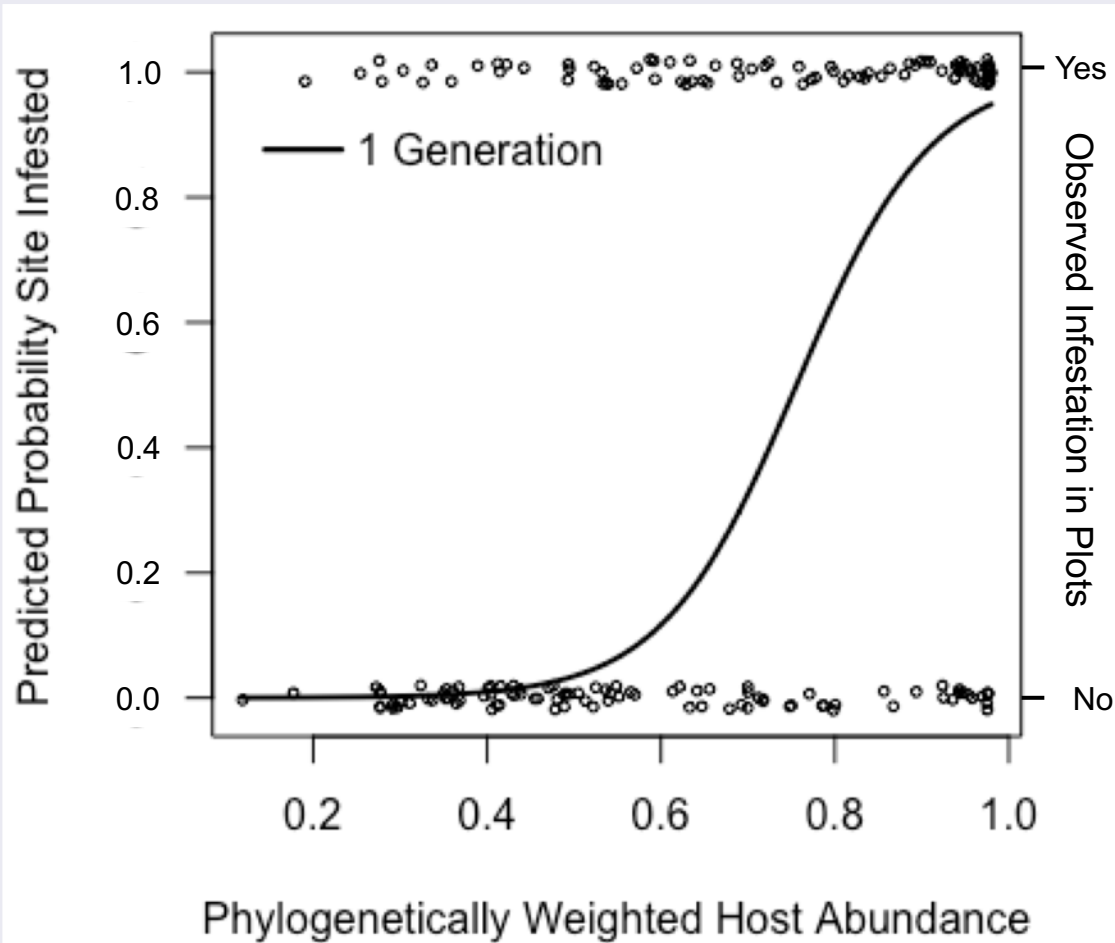
Temp (C)



gens



Temp (C)

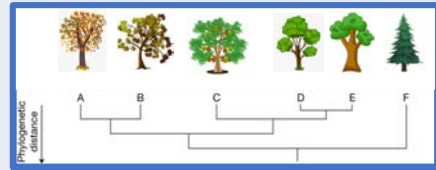


$$p(\text{Site Suscept}) = 16.9 * wpS + 3.1 * Gen - 3.9 * (wpS * Gen) - 12.9$$

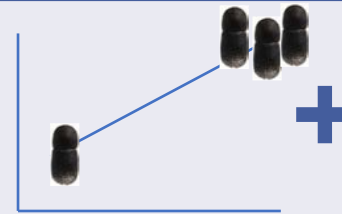
Community Structure and Climate Effects



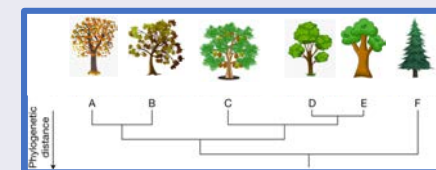
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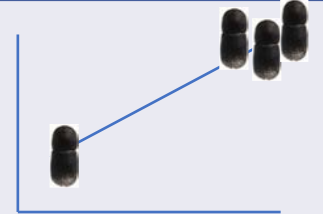
gens



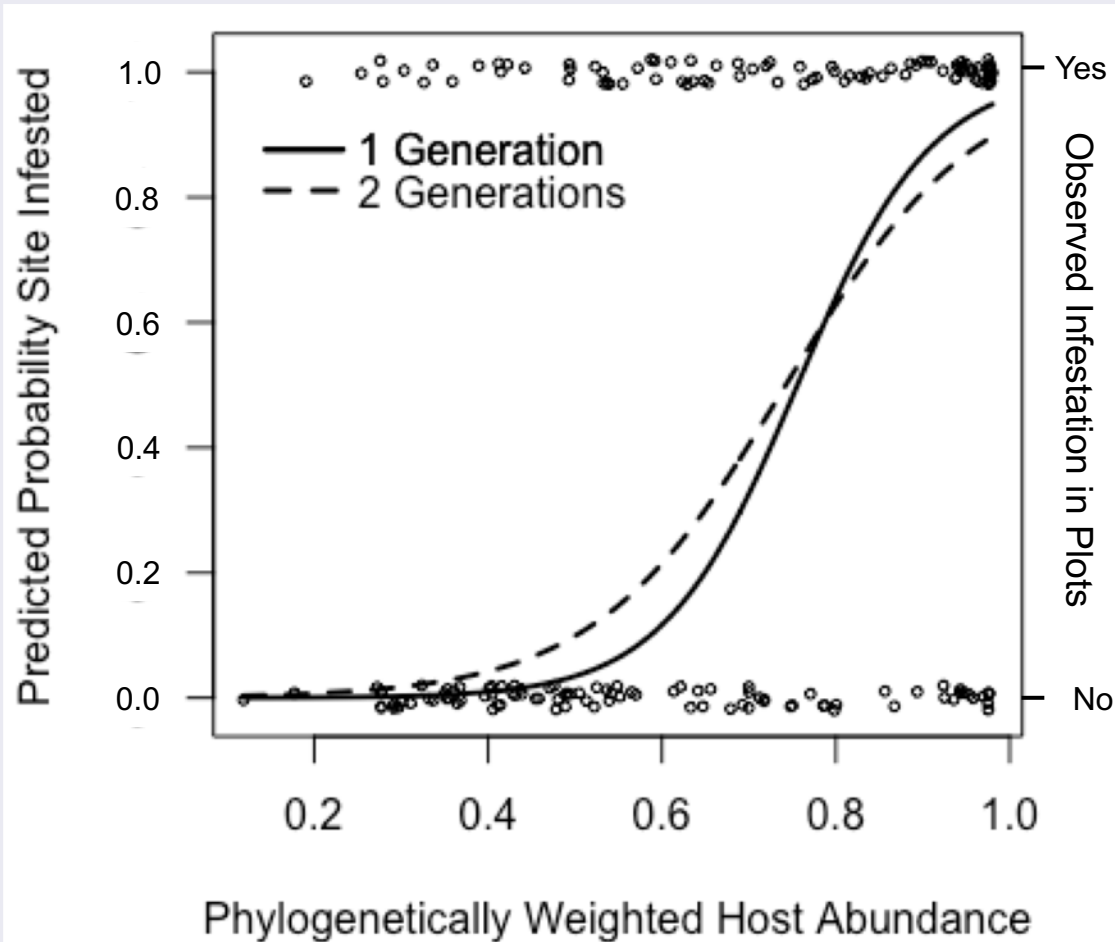
Temp (C)



gens



Temp (C)

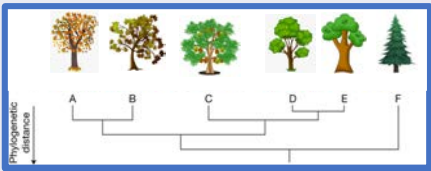


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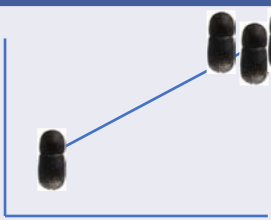
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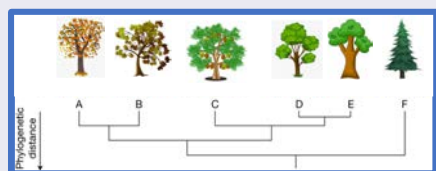
$p(\text{Site Suscept}) =$



$+$ # gens

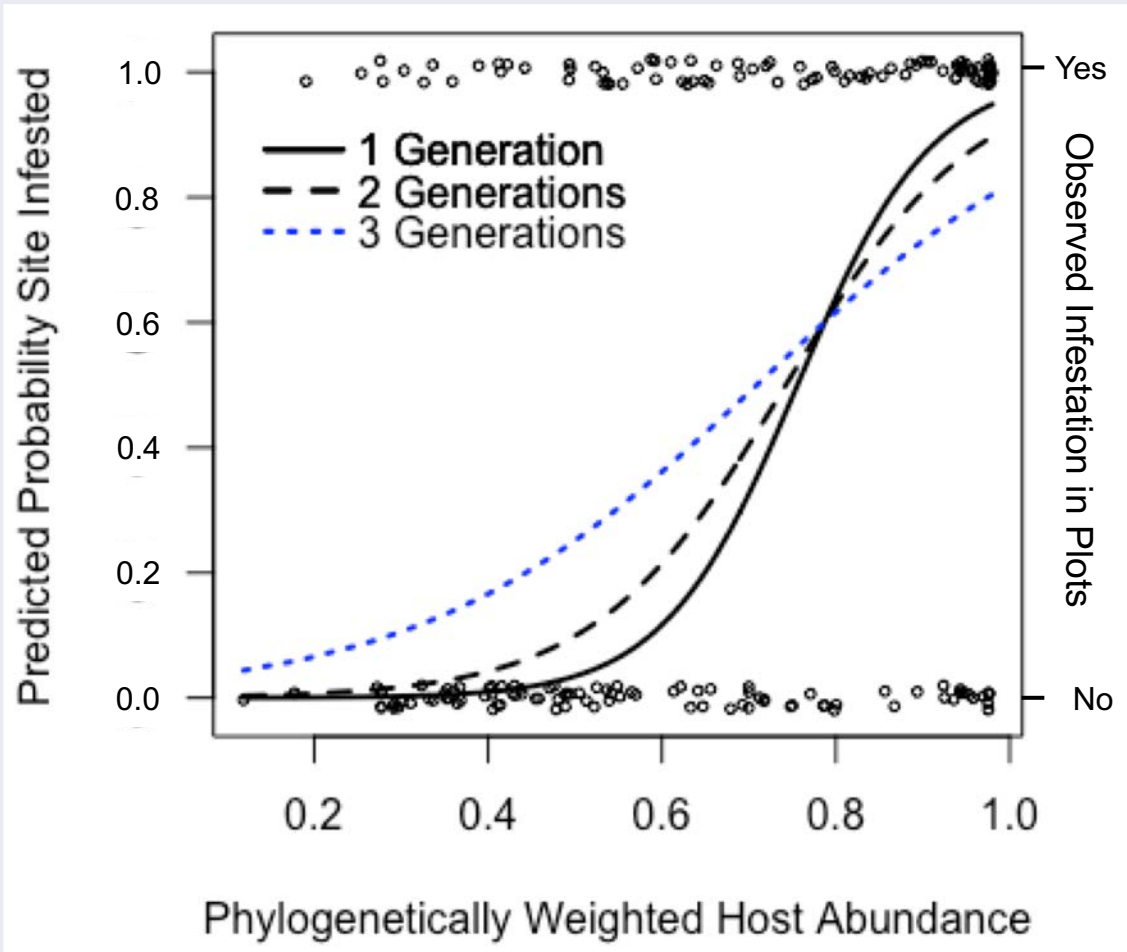


$+$



$*$ # gens

Temp (C)



Warmer Conditions = More beetle generations

Unfavorable communities are more susceptible

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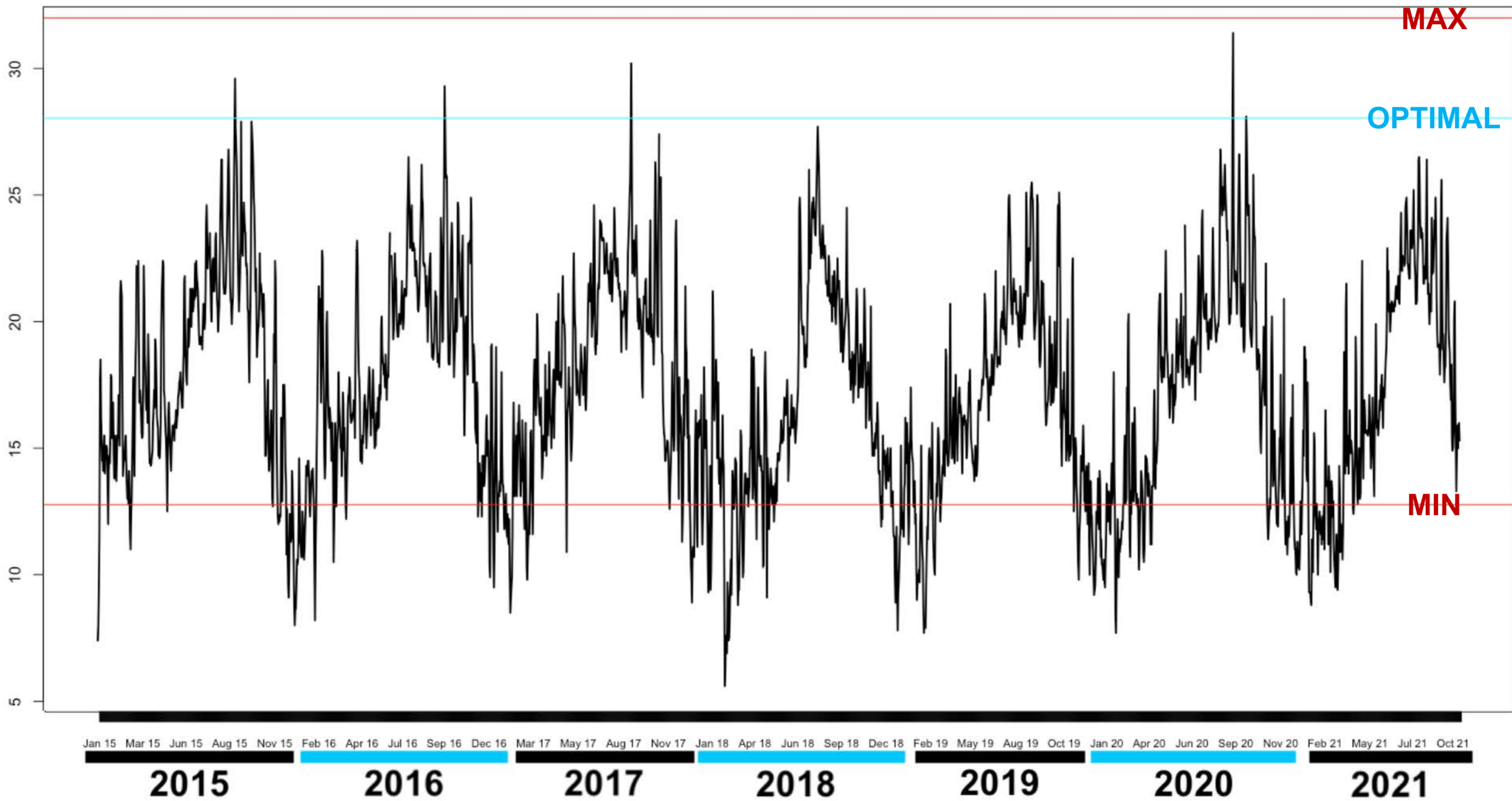


Warmer Conditions = More beetle generations

Unfavorable communities are more susceptible

- ❖ Focus monitoring resources in non-infested locations with high likelihoods of being infested
- ❖ Prioritize management actions in infested locations where community composition AND microclimate are most favorable for beetle establishment

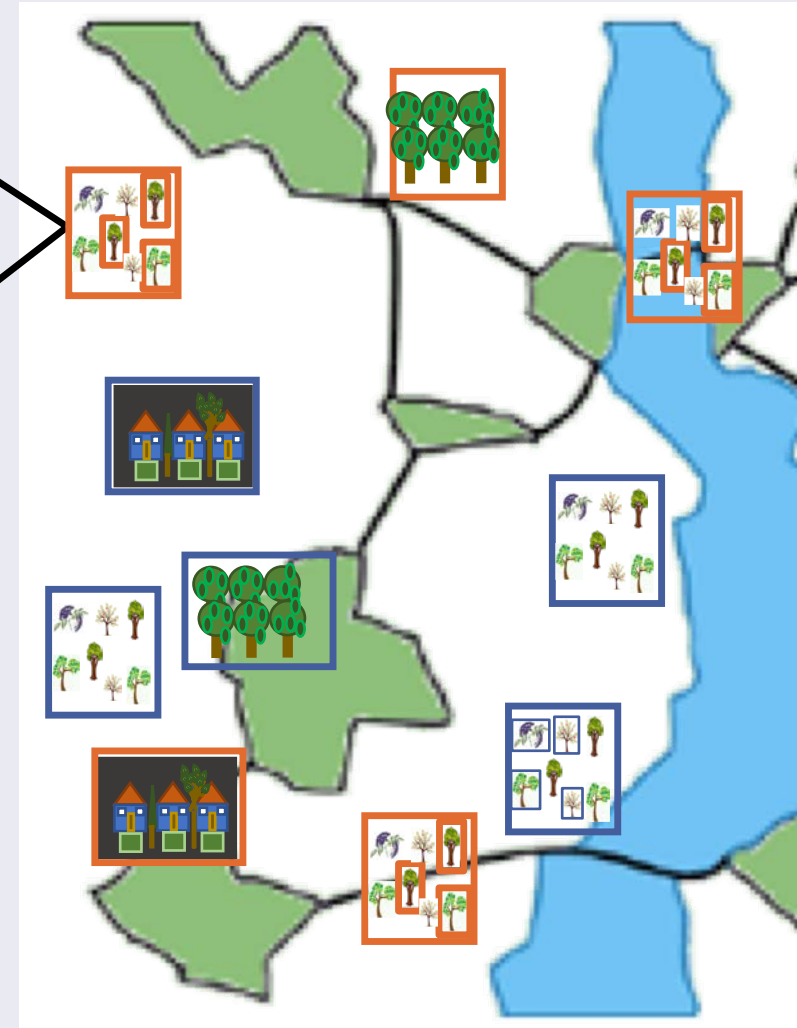
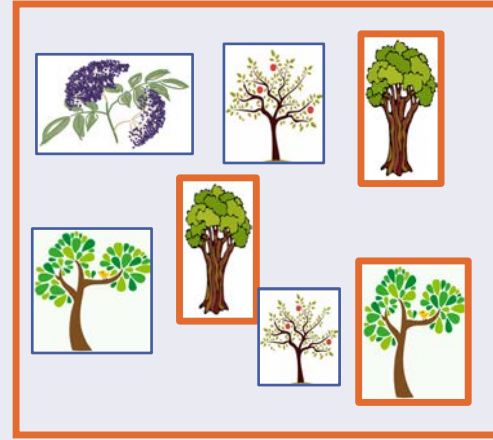
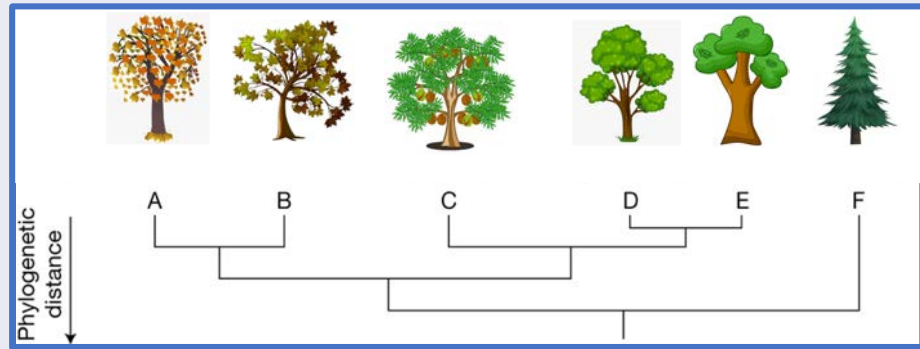
Temp (C)



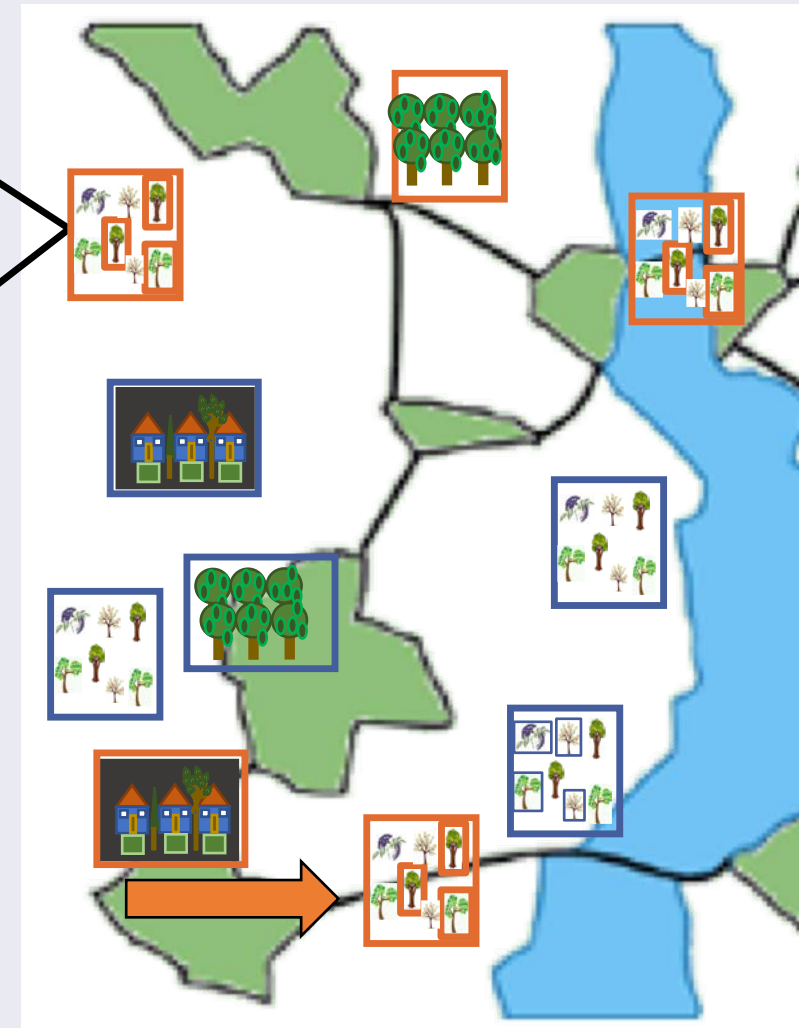
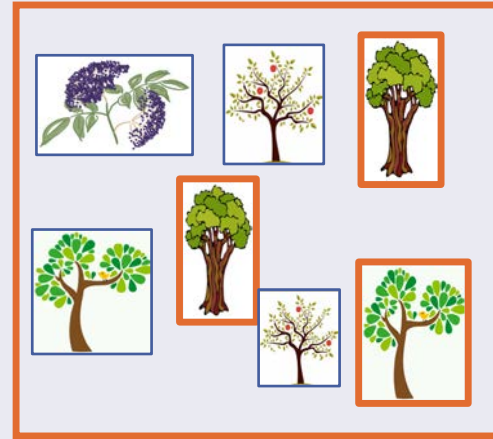
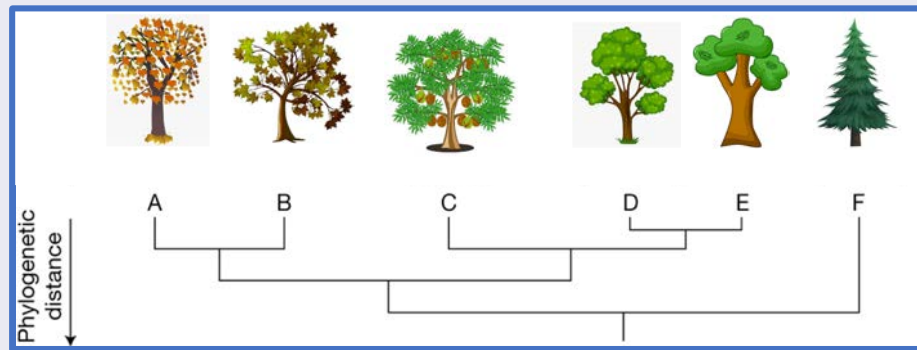


- Statewide predictions over time
- Landscape considerations

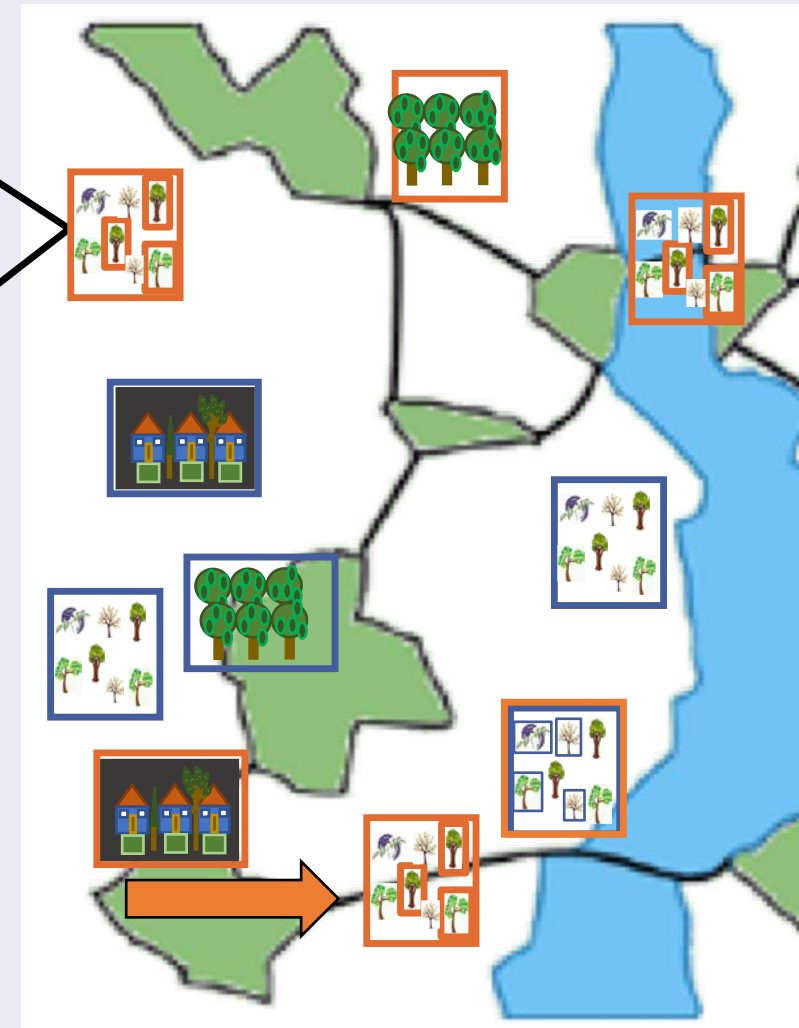
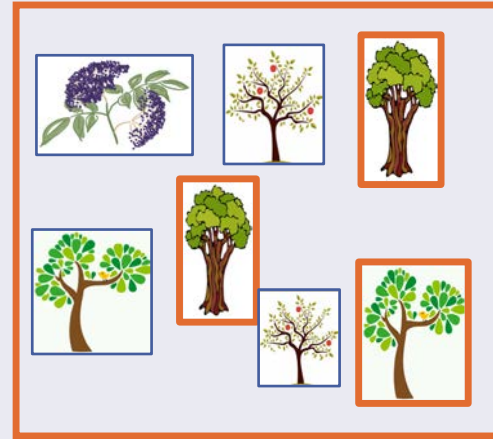
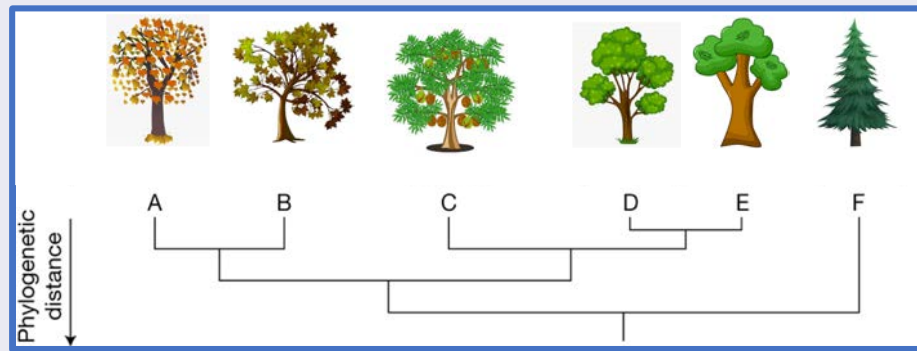
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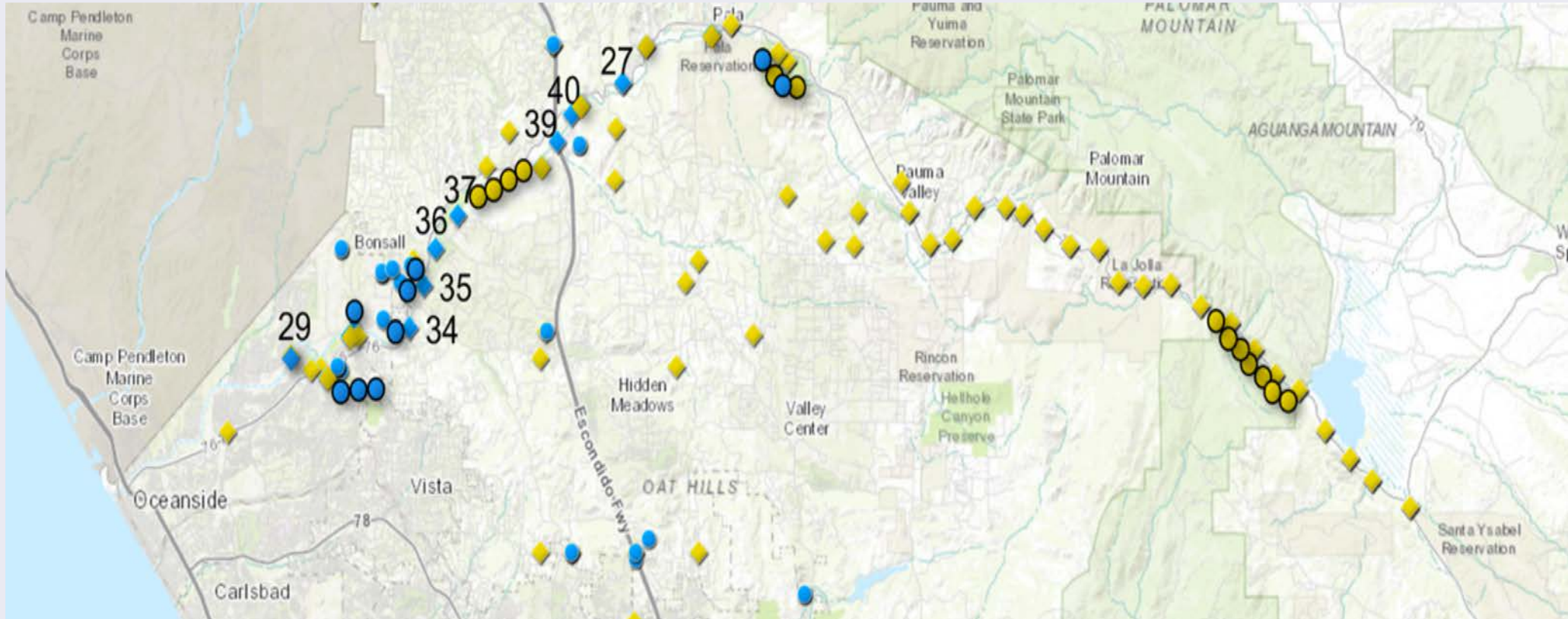
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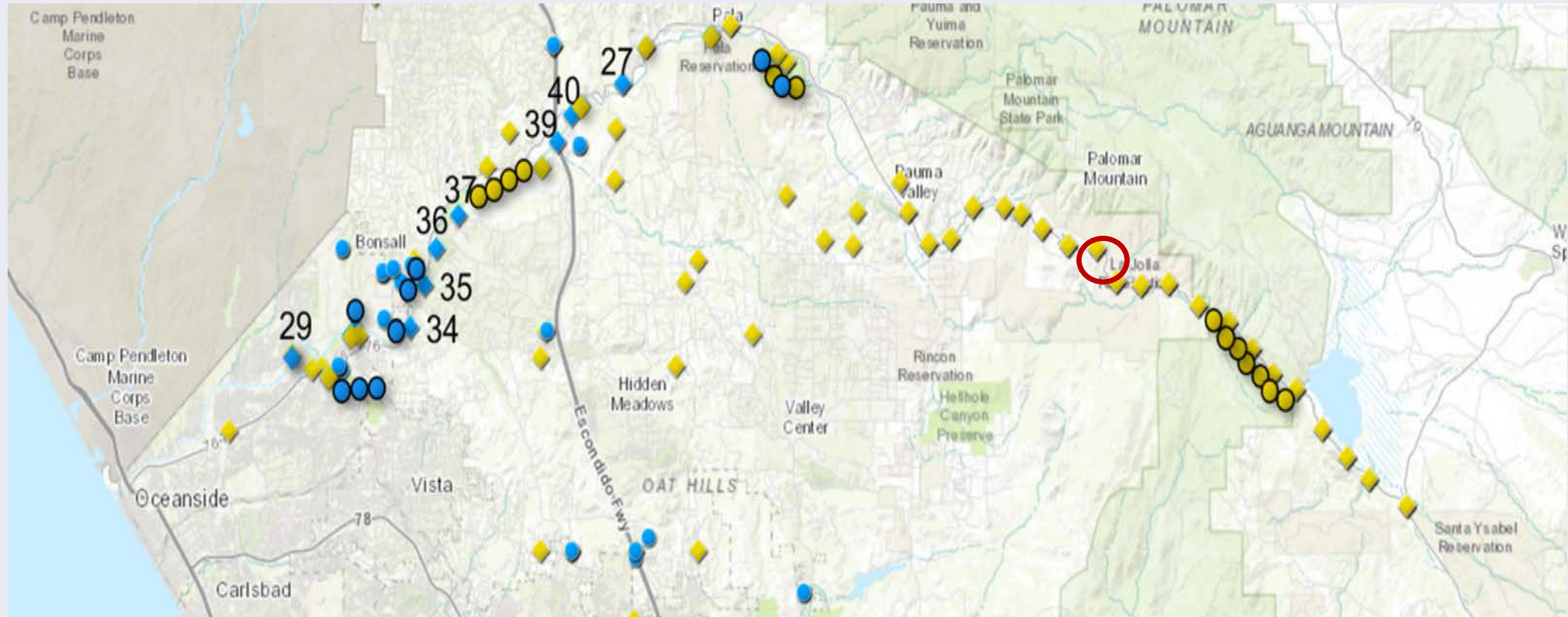


Monitoring beetle population in San Luis Rey River



San Luis Rey monitoring traps and permanent plot locations. Blue circles are KSHB positive traps, yellow diamonds are negative monitoring traps. The red circle is recent findings

Monitoring beetle population in San Luis Rey River



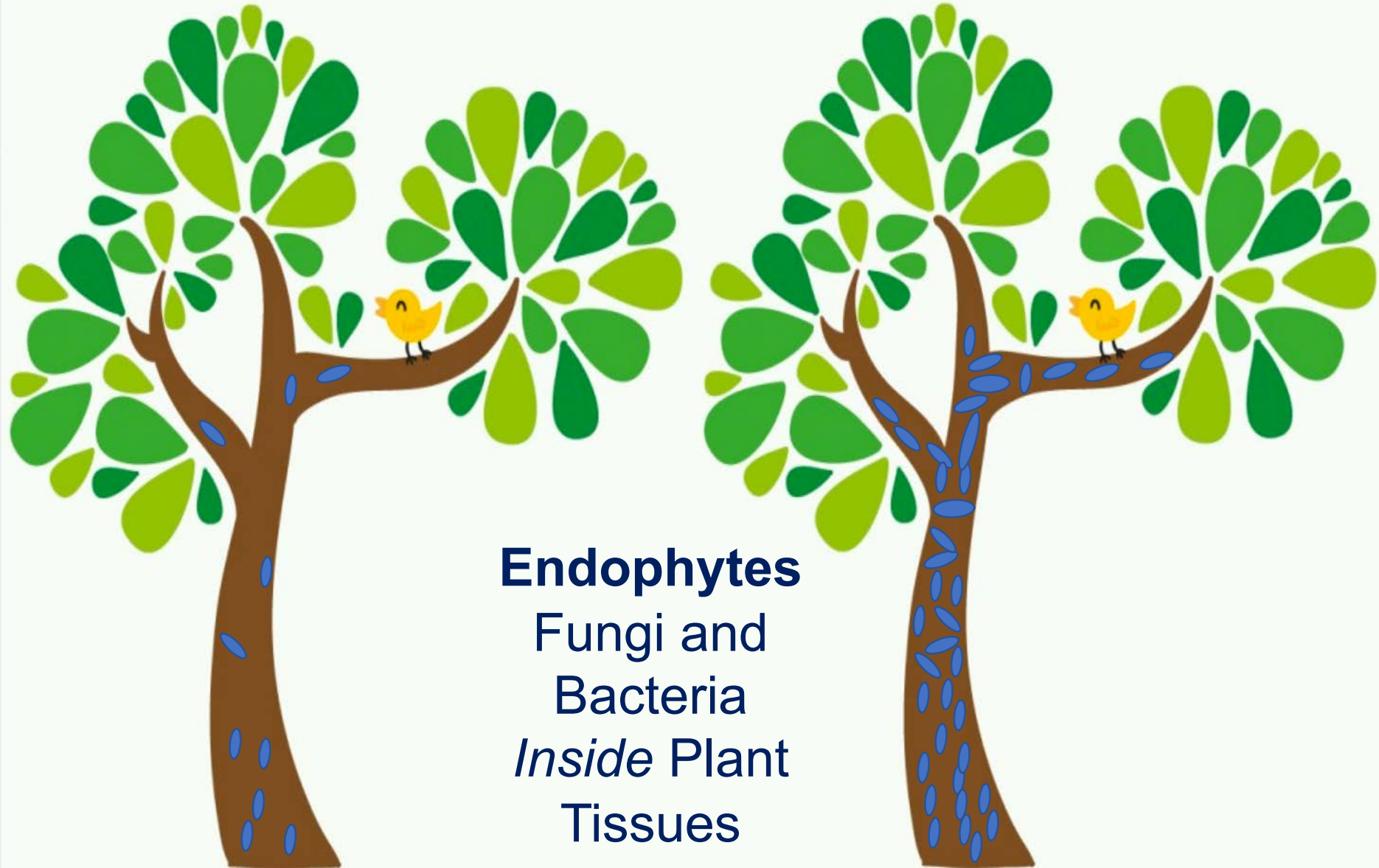
San Luis Rey monitoring traps and permanent plot locations. Blue circles are KSHB positive traps, yellow diamonds are negative monitoring traps. The red circle is recent findings

**New KSHB infestation on a sycamore (*Platanus racemosa*) along SLR near Lilac Rd.
No new infestation was observed on any willows and cottonwoods in the area.**



New PSHB infestation on a sycamore (*Platanus racemosa*) along SLR near Oak Knoll Campground





Endophytes
Fungi and
Bacteria
Inside Plant
Tissues

Non-Infested Sycamore in a Disease Hot-Spot



March 2016



November 2016

Endophyte Sampling



- Willows
- Cottonwood
- Oak
- Sycamore



Total 606 samples were collected in San Diego

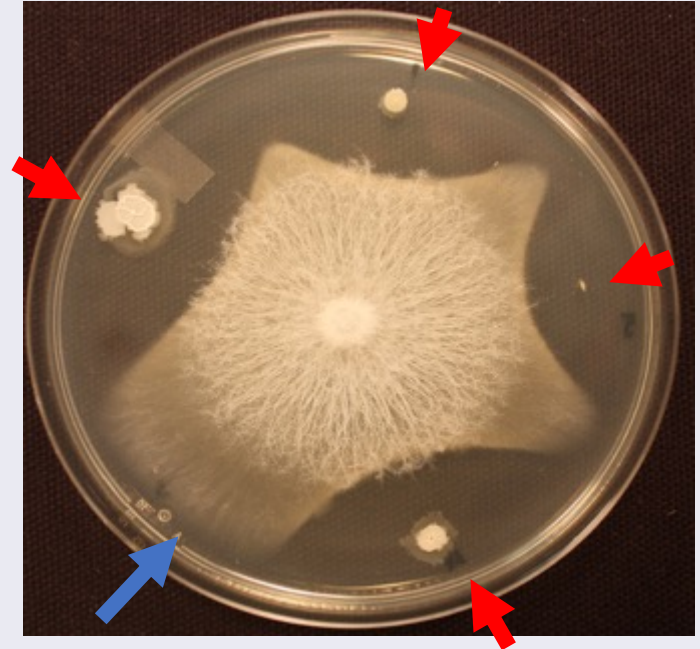
Preliminary Endophyte Screening



In vitro Inhibition Bioassays



Control



Treatment

● No Inhibition

● Inhibition

Microbes Exhibiting Inhibition of *Fusarium* growth

Bacterial Inhibition

Pseudomonas sp.

Pantoea sp.

Variovorax sp.

Bacillus spp.

Fungal Inhibition

*Aureobasidium pullulans**

Pithomyces chartarum

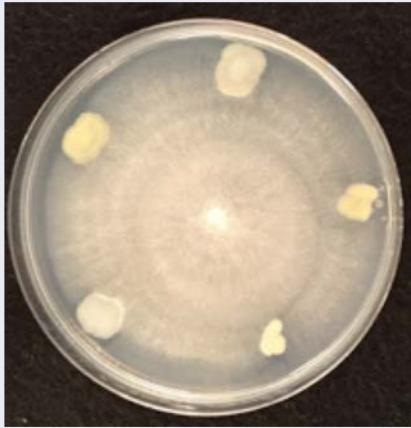
Acremonium sp.

Alternaria alternate

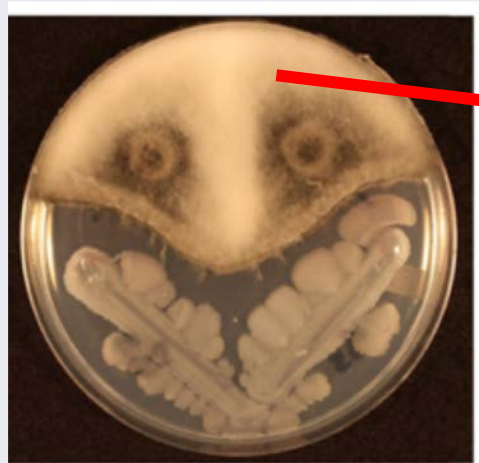
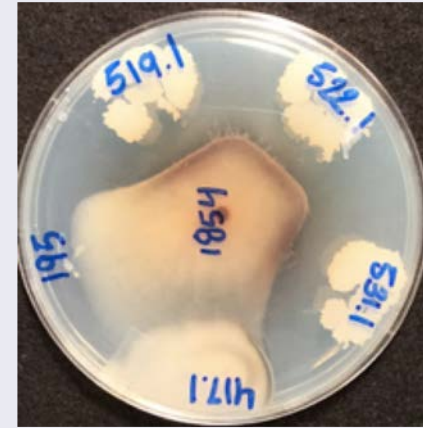
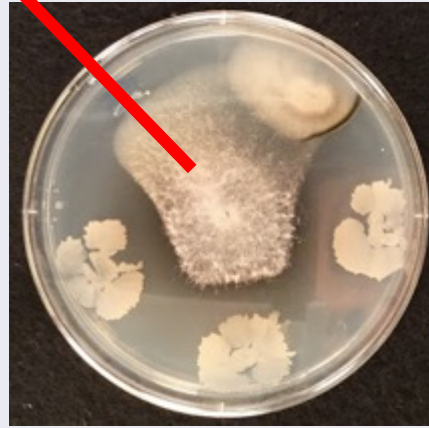
Epicocoum nigrum

Endophyte Sampling

Fusarium kuroshium

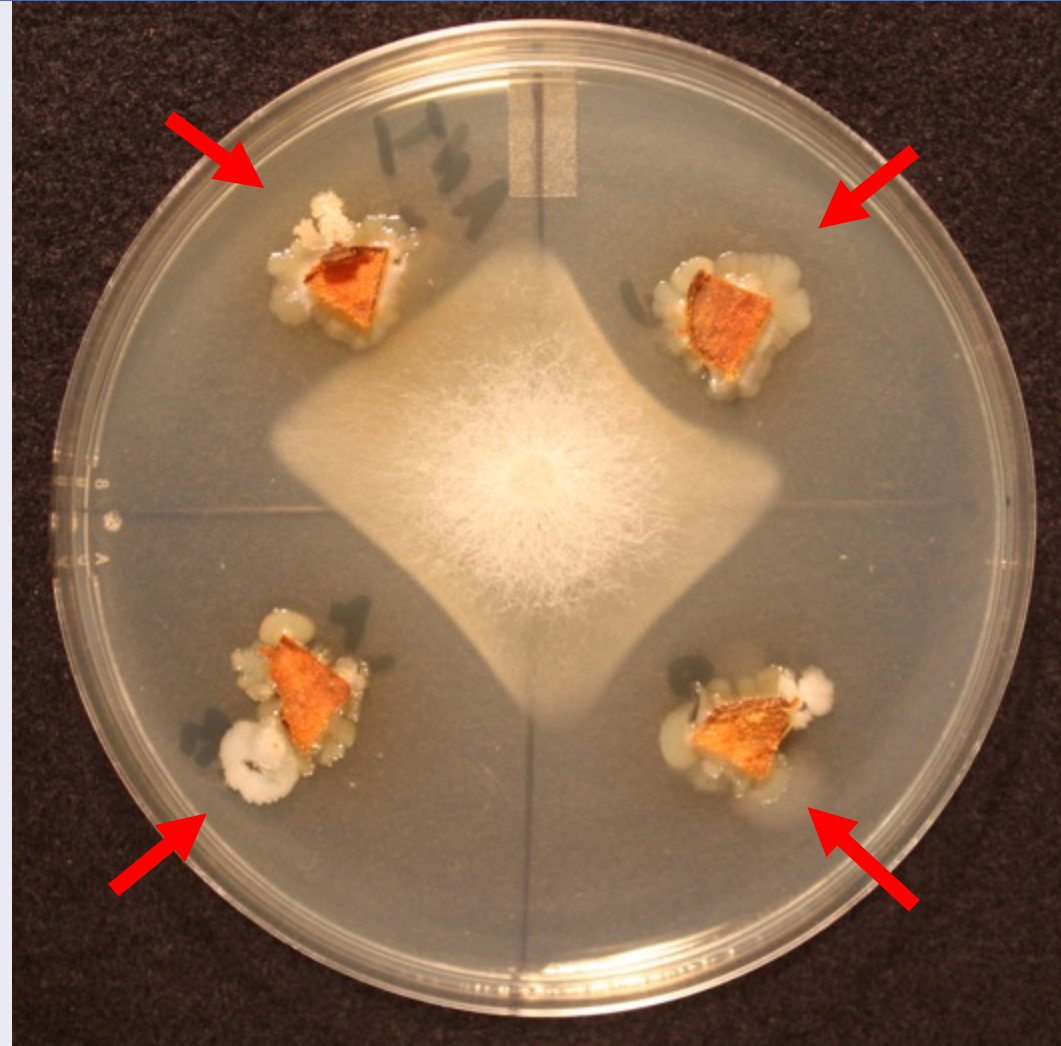


Control



Botryosphaeria sp.

Inhibition Bioassays

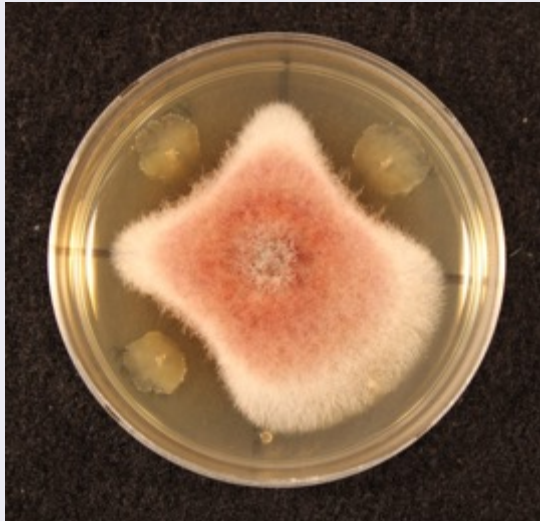


In planta

● No Inhibition ● Inhibition

Restoration with Biocontrol

Recover



Willow cutting for propagation

Infiltrate

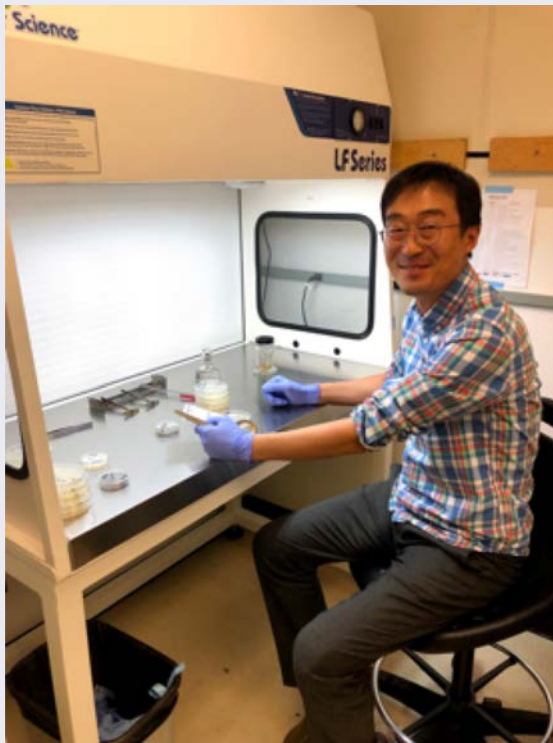


Propagate



Restore

Fermentation of endophytic bacteria in large scale



In vitro isolation and culturing



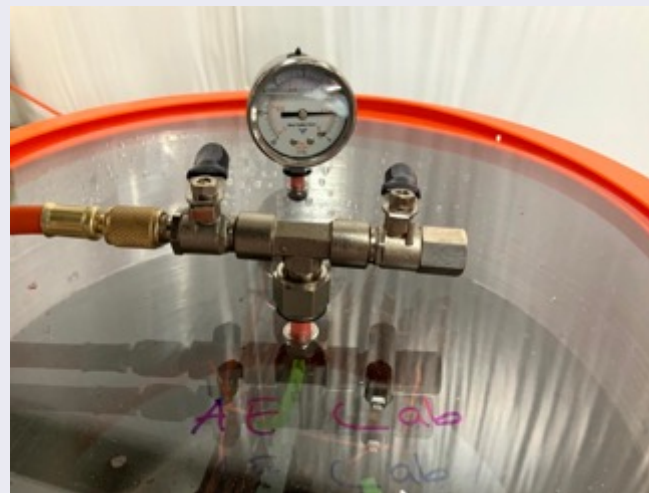
In vitro fermentation



Large scale fermentation

Collaboration with a group of scientists from the University of Chonnam from South Korea on fermentation of endophytic bacteria that could be applied in large scale.

Delivering endophytes into propagation cuttings via vacuum infiltration



Acknowledgement



Eskalen Lab, UC Davis
Richard Stouthamer, UC Riverside
Paul Rugman-Jones, UC Riverside
John Kabashima, UCCE Orange County
Milan Mitrovich NCC, Orange County
Kristine Preston, USGS
Kim Smith, SANDAG
Zack, The Nature Conservancy
Ben Faber, Farm Advisor, Ventura County
Richard Demerjian, UC Irvine
Dan Berry, Huntington Botanical Garden
Jim Folsom, Huntington Botanical Garden

Mary Lu Arpaia, UC Riverside
Kim Corella, Cal Fire
Tom Smith, Cal Fire
Linda Bellamy, Ventura
Susan Frankel, USDA Forest Service
Tom Atkinson, University of Texas
Jim Downer, UCCE Ventura
Faith Campbell, Center for Invasive Species



Sea & Sage Audubon Society
An Orange County Chapter of the National Audubon Society

